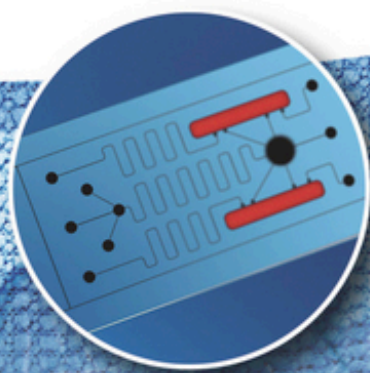


Liang-Yin Chu and Wei Wang

# Microfluidics for Advanced Functional Polymeric Materials





**Microfluidics for Advanced  
Functional Polymeric Materials**



# **Microfluidics for Advanced Functional Polymeric Materials**

*Liang-Yin Chu and Wei Wang*

**WILEY-VCH**

## Authors

### *Professor Liang-Yin Chu*

Sichuan University  
School of Chemical Engineering  
No. 24, Yihuan Road  
First Southern Section  
610065 Chengdu  
China

### *Dr. Wei Wang*

Sichuan University  
School of Chemical Engineering  
No. 24 Yihuan Road  
First Southern Section  
610065 Chengdu  
China

## Cover

Microchip – fotolia\_@science photo;  
Microfiber structure – fotolia\_@nelik;  
Nanostructure – fotolia\_@chaoss

All books published by **Wiley-VCH** are carefully produced. Nevertheless, authors, editors, and publisher do not warrant the information contained in these books, including this book, to be free of errors. Readers are advised to keep in mind that statements, data, illustrations, procedural details or other items may inadvertently be inaccurate.

**Library of Congress Card No.:** applied for

### **British Library Cataloguing-in-Publication Data**

A catalogue record for this book is available from the British Library.

### **Bibliographic information published by the Deutsche Nationalbibliothek**

The Deutsche Nationalbibliothek lists this publication in the Deutsche Nationalbibliografie; detailed bibliographic data are available on the Internet at <<http://dnb.d-nb.de>>.

© 2017 Wiley-VCH Verlag GmbH & Co. KGaA, Boschstr. 12, 69469 Weinheim, Germany

All rights reserved (including those of translation into other languages). No part of this book may be reproduced in any form – by photoprinting, microfilm, or any other means – nor transmitted or translated into a machine language without written permission from the publishers. Registered names, trademarks, etc. used in this book, even when not specifically marked as such, are not to be considered unprotected by law.

**Print ISBN:** 978-3-527-34182-5

**ePDF ISBN:** 978-3-527-80366-8

**ePub ISBN:** 978-3-527-80365-1

**Mobi ISBN:** 978-3-527-80364-4

**oBook ISBN:** 978-3-527-80363-7

**Cover Design** Adam-Design, Weinheim, Germany

**Typesetting** SPi Global, Chennai, India

**Printing and Binding**

Printed on acid-free paper

## Contents

### Preface *xiii*

- 1 Introduction 1**
- 1.1 Microfluidics and Its Superiority in Controllable Fabrication of Functional Materials 1
- 1.2 Microfluidic Fabrication of Microspheres and Microcapsules from Microscale Closed Liquid–Liquid Interfaces 3
- 1.3 Microfluidic Fabrication of Membranes in Microchannels from Microscale Nonclosed Layered Laminar Interfaces 4
- 1.4 Microfluidic Fabrication of Microfiber Materials from Microscale Nonclosed Annular Laminar Interfaces 5
- References 6
  
- 2 Shear-Induced Generation of Controllable Multiple Emulsions in Microfluidic Devices 11**
- 2.1 Introduction 11
- 2.2 Microfluidic Strategy for Shear-Induced Generation of Controllable Emulsion Droplets 12
- 2.3 Shear-Induced Generation of Controllable Monodisperse Single Emulsions 14
- 2.4 Shear-Induced Generation of Controllable Multiple Emulsions 16
- 2.4.1 Shear-Induced Generation of Controllable Double Emulsions 16
- 2.4.2 Shear-Induced Generation of Controllable Triple Emulsions 19
- 2.5 Shear-Induced Generation of Controllable Multicomponent Multiple Emulsions 22
- 2.5.1 Shear-Induced Generation of Controllable Quadruple-Component Double Emulsions 22
- 2.5.2 Extended Microfluidic Device for Controllable Generation of More Complex Multicomponent Multiple Emulsions 27
- 2.6 Summary 31
- References 31

<b>3</b>	<b>Wetting-Induced Generation of Controllable Multiple Emulsions in Microfluidic Devices</b>	<b>35</b>
3.1	Introduction	35
3.2	Microfluidic Strategy for Wetting-Induced Production of Controllable Emulsions	36
3.2.1	Strategy for Wetting-Induced Production of Controllable Emulsions via Wetting-Induced Spreading	36
3.2.2	Strategy for Wetting-Induced Production of Controllable Emulsions via Wetting-Induced Coalescing	37
3.3	Generation of Controllable Multiple Emulsions via Wetting-Induced Spreading	38
3.3.1	Wetting-Induced Generation of Monodisperse Controllable Double Emulsions	38
3.3.2	Wetting-Induced Generation of Monodisperse Higher Order Multiple Emulsions	41
3.3.3	Wetting-Induced Generation of Monodisperse Multiple Emulsions via Droplet-Triggered Droplet Pairing	44
3.4	Generation of Controllable Multiple Emulsions via Wetting-Induced Droplet Coalescing	47
3.5	Summary	50
	References	52
<b>4</b>	<b>Microfluidic Fabrication of Monodisperse Hydrogel Microparticles</b>	<b>55</b>
4.1	Introduction	55
4.2	Microfluidic Fabrication of Monodisperse PNIPAM Hydrogel Microparticles for Sensing Tannic Acid (TA)	55
4.2.1	Microfluidic Fabrication of Monodisperse PNIPAM Hydrogel Microparticles	56
4.2.2	Volume-Phase Transition Behaviors of PNIPAM Microgels Induced by TA	57
4.3	Microfluidic Fabrication of Monodisperse Core–Shell PNIPAM Hydrogel Microparticles for Sensing Ethyl Gallate (EG)	62
4.3.1	Microfluidic Fabrication of Monodisperse Core–Shell PNIPAM Hydrogel Microparticles	62
4.3.2	Thermo-Responsive Phase Transition Behaviors of PNIPAM Microspheres in EG Solution	65
4.3.3	The Intact-to-Broken Transformation Behaviors of Core–Shell PNIPAM Microcapsules in Aqueous Solution with Varying EG Concentrations	65
4.4	Microfluidic Fabrication of Monodisperse Core–Shell Hydrogel Microparticles for the Adsorption and Separation of Pb <sup>2+</sup>	67
4.4.1	Microfluidic Fabrication of Monodisperse Core–Shell Microparticles with Magnetic Core and Hydrogel Shell	68



- 4.4.2 Pb<sup>2+</sup> Adsorption Behaviors of Magnetic PNB Core–Shell  
Microspheres 71
- 4.5 Summary 75
- References 76
  
- 5 Microfluidic Fabrication of Monodisperse Porous  
Microparticles 79**
- 5.1 Introduction 79
- 5.2 Microfluidic Fabrication of Monodisperse Porous Poly(HEMA-MMA)  
Microparticles 79
  - 5.2.1 Microfluidic Fabrication Strategy 80
  - 5.2.2 Structures of Poly(HEMA-MMA) Porous Microspheres 82
- 5.3 Microfluidic Fabrication of Porous PNIPAM Microparticles with  
Tunable Response Behaviors 83
  - 5.3.1 Microfluidic Fabrication Strategy 86
  - 5.3.2 Tunable Response Behaviors of Porous PNIPAM Microparticles 87
- 5.4 Microfluidic Fabrication of PNIPAM Microparticles with Open-Celled  
Porous Structure for Fast Response 90
  - 5.4.1 Microfluidic Fabrication Strategy 91
  - 5.4.2 Morphologies and Microstructures of Porous PNIPAM  
Microparticles 93
  - 5.4.3 Thermo-Responsive Volume Change Behaviors of PNIPAM Porous  
Microparticles 98
- 5.5 Summary 103
- References 103
  
- 6 Microfluidic Fabrication of Uniform Hierarchical Porous  
Microparticles 105**
- 6.1 Introduction 105
- 6.2 Microfluidic Strategy for Fabrication of Uniform Hierarchical Porous  
Microparticles 106
- 6.3 Controllable Microfluidic Fabrication of Uniform Hierarchical Porous  
Microparticles 108
  - 6.3.1 Preparation of Hierarchical Porous Microparticles 108
  - 6.3.2 Hierarchical Porous Microparticles with Micrometer-Sized Pores from  
Deformed W/O/W Emulsions 108
  - 6.3.3 Integration of Nanometer- and Micrometer-Sized Pores for Creating  
Hierarchical Porous Microparticles 111
- 6.4 Hierarchical Porous Microparticles for Oil Removal 114
  - 6.4.1 Concept of the Hierarchical Porous Microparticles for Oil  
Removal 114
  - 6.4.2 Hierarchical Porous Microparticles for Magnetic-Guided Oil  
Removal 115
- 6.5 Hierarchical Porous Microparticles for Protein Adsorption 116

- 6.5.1 Concept of Hierarchical Porous Microparticles for Protein Adsorption 116
- 6.5.2 Hierarchical Porous Microparticles for Enhanced Protein Adsorption 117
- 6.6 Summary 118  
References 118
  
- 7 Microfluidic Fabrication of Monodisperse Hollow Microcapsules 123**
- 7.1 Introduction 123
- 7.2 Microfluidic Fabrication of Monodisperse Ethyl Cellulose Hollow Microcapsules 124
  - 7.2.1 Microfluidic Fabrication Strategy 124
  - 7.2.2 Morphologies and Structures of Ethyl Cellulose Hollow Microcapsules 125
- 7.3 Microfluidic Fabrication of Monodisperse Calcium Alginate Hollow Microcapsules 131
  - 7.3.1 Microfluidic Fabrication Strategy 132
  - 7.3.2 Morphologies and Structures of Calcium Alginate Hollow Microcapsules 133
- 7.4 Microfluidic Fabrication of Monodisperse Glucose-Responsive Hollow Microcapsules 136
  - 7.4.1 Microfluidic Fabrication Strategy 136
  - 7.4.2 Glucose-Responsive Behaviors of Microcapsules 140
  - 7.4.3 Glucose-Responsive Drug Release Behaviors of Microcapsules 142
- 7.5 Microfluidic Fabrication of Monodisperse Multi-Stimuli-Responsive Hollow Microcapsules 144
  - 7.5.1 Microfluidic Fabrication Strategy 145
  - 7.5.2 Stimuli-Responsive Behaviors of Microcapsules 150
  - 7.5.3 Controlled-Release Characteristics of Multi-Stimuli-Responsive Microcapsules 154
- 7.6 Summary 158  
References 158
  
- 8 Microfluidic Fabrication of Monodisperse Core-Shell Microcapsules 161**
- 8.1 Introduction 161
- 8.2 Microfluidic Strategy for Fabrication of Monodisperse Core-Shell Microcapsules 162
- 8.3 Smart Core-Shell Microcapsules for Thermo-Triggered Burst Release 162
  - 8.3.1 Fabrication of Core-Shell Microcapsules for Thermo-Triggered Burst Release of Oil-Soluble Substances 162
  - 8.3.2 Fabrication of Core-Shell Microcapsules for Thermo-Triggered Burst Release of Nanoparticles 166
  - 8.3.3 Fabrication of Core-Shell Microcapsules for Direction-Specific Thermo-Responsive Burst Release 168

- 8.4 Smart Core–Shell Microcapsules for Alcohol-Responsive Burst Release 171
- 8.5 Smart Core–Shell Microcapsules for K<sup>+</sup>-Responsive Burst Release 174
- 8.6 Smart Core–Shell Microcapsules for pH-Responsive Burst Release 176
  - 8.6.1 Concept of the Core–Shell Microcapsules for pH-Responsive Burst Release 176
  - 8.6.2 Fabrication of the Core–Shell Chitosan Microcapsules 177
  - 8.6.3 Core–Shell Chitosan Microcapsules for pH-Responsive Burst Release 179
- 8.7 Summary 182
  - References 183
- 9 Microfluidic Fabrication of Monodisperse Hole–Shell Microparticles 187**
  - 9.1 Introduction 187
  - 9.2 Microfluidic Strategy for Fabrication of Monodisperse Hole–Shell Microparticles 188
  - 9.3 Hole–Shell Microparticles for Thermo-Driven Crawling Movement 188
    - 9.3.1 Concept of the Hole–Shell Microparticles for Thermo-Driven Crawling Movement 188
    - 9.3.2 Fabrication of Hole–Shell Microparticles for Thermo-Driven Crawling Movement 190
    - 9.3.3 Effect of Inner Cavity on the Thermo-Responsive Volume-Phase Transition Behaviors of Hole–Shell Microparticles 191
    - 9.3.4 Hole–Shell Microparticles for Thermo-Driven Crawling Movement 193
  - 9.4 Hole–Shell Microparticles for Pb<sup>2+</sup> Sensing and Actuating 195
    - 9.4.1 Fabrication of Hole–Shell Microparticles for Pb<sup>2+</sup> Sensing and Actuating 195
    - 9.4.2 Magnetic-Guided Targeting Behavior of Poly(NIPAM-*co*-B18C6Am) Hole–Shell Microparticles 195
    - 9.4.3 Effects of Pb<sup>2+</sup> on the Thermo-Responsive Volume Change Behaviors of Poly(NIPAM-*co*-B18C6Am) Hole–Shell Microparticles 196
    - 9.4.4 Effects of Hollow Cavity on the Time-Dependent Volume Change Behaviors of Poly(NIPAM-*co*-B18C6Am) Hole–Shell Microparticles 199
    - 9.4.5 Micromanipulation of Poly(NIPAM-*co*-B18C6Am) Hole–Shell Microparticles for Preventing Pb<sup>2+</sup> Leakage from Microcapillary 200
  - 9.5 Hole–Shell Microparticles for Controlled Capture and Confined Microreaction 201
    - 9.5.1 Microfluidic Fabrication of Hole–Shell Microparticles 201
    - 9.5.2 Precise Control over the Hole–Shell Structure of the Microparticles 203
    - 9.5.3 Precise Control over the Functionality of Hollow Core Surface 205

- 9.5.4 Hole–Shell Microparticles for Controlled Capture and Confined Microreaction 206
- 9.6 Summary 207  
References 207
  
- 10 Microfluidic Fabrication of Controllable Multicompartmental Microparticles 211**
  - 10.1 Introduction 211
  - 10.2 Microfluidic Strategy for the Fabrication of Controllable Multicompartmental Microparticles 212
  - 10.3 Multi-core/Shell Microparticles for Co-encapsulation and Synergistic Release 212
    - 10.3.1 Microfluidic Fabrication of Multi-core/Shell Microparticles 212
    - 10.3.2 Multi-core/Shell Microparticles for Controllable Co-encapsulation 213
    - 10.3.3 Multi-core/Shell Microparticles for Synergistic Release 216
  - 10.4 Trojan-Horse-Like Microparticles for Co-delivery and Programmed Release 217
    - 10.4.1 Fabrication of Trojan-Horse-Like Microparticles from Triple Emulsions 217
  - 10.5 Summary 218  
References 219
  
- 11 Microfluidic Fabrication of Functional Microfibers with Controllable Internals 223**
  - 11.1 Introduction 223
  - 11.2 Microfluidic Strategy for Fabrication of Functional Microfibers with Controllable Internals 224
  - 11.3 Core–Sheath Microfibers with Tubular Internals for Encapsulation of Phase Change Materials 224
    - 11.3.1 Fabrication of Core–Sheath Microfibers with Tubular Internals 224
    - 11.3.2 Morphological Characterization of the Core–Sheath Microfibers 227
    - 11.3.3 Thermal Property of the Core–Sheath Microfibers 227
    - 11.3.4 Core–Sheath Microfibers for Temperature Regulation 230
  - 11.4 Peapod-Like Microfibers with Multicompartmental Internals for Synergistic Encapsulation 235
    - 11.4.1 Fabrication of Peapod-Like Microfibers with Multicompartmental Internals 236
    - 11.4.2 Effects of Flow Rates on the Structures of Peapod-Like Jet Templates and Chitosan Microfibers 236
    - 11.4.3 Peapod-Like Chitosan Microfibers with Multicompartment Internals for Synergistic Encapsulation 240
  - 11.5 Spider-Silk-Like Microfibers with Spindle-Knot Internals for 3D Assembly and Water Collection 241
    - 11.5.1 Fabrication of Spider-Silk-Like Microfibers with Spindle-Knot Internals 241

- 11.5.2 Morphological Characterization of the Jet Templates and Spider-Silk-Like Microfibers 242
- 11.5.3 Magnetic-Guided Patterning and Assembling of the Ca-Alginate Microfibers 244
- 11.5.4 Water Collection of Dehydrated Ca-Alginate Microfibers with Magnetic Spindle-Knot Internals 246
- 11.6 Summary 248
- References 248
  
- 12 Microfluidic Fabrication of Membrane-in-a-Chip with Self-Regulated Permeability 253**
- 12.1 Introduction 253
- 12.2 Microfluidic Strategy for Fabrication of Membrane-in-a-Chip 254
- 12.3 Temperature- and Ethanol-Responsive Smart Membrane in Microchip for Detection 255
- 12.3.1 Fabrication of Nanogel-Containing Smart Membrane in Microchip 255
- 12.3.2 Temperature-Responsive Self-Regulation of the Membrane Permeability 257
- 12.3.3 Ethanol-Responsive Self-Regulation of the Membrane Permeability 260
- 12.3.4 Reversible and Repeated Thermo/Ethanol-Responsive Self-Regulation of the Membrane Permeability 263
- 12.4 Summary 264
- References 264
  
- 13 Microfluidic Fabrication of Microvalve-in-a-Chip 267**
- 13.1 Introduction 267
- 13.2 Microfluidic Strategy for Fabrication of Microvalve-in-a-Chip 268
- 13.2.1 Fabrication of Thermo-Responsive Hydrogel Microvalve within Microchip for Thermostatic Control 268
- 13.2.2 Fabrication of Pb<sup>2+</sup>-Responsive Hydrogel Microvalve within Microchip for Pb<sup>2+</sup> Detection 270
- 13.3 Smart Microvalve-in-a-Chip with Thermostatic Control for Cell Culture 272
- 13.3.1 Setup of Microvalve-Integrated Micro-heat-Exchanging System 273
- 13.3.2 Thermo-Responsive Switch Performance of Hydrogel Microvalve 274
- 13.3.3 Sealing Performance of Hydrogel Microvalve 276
- 13.3.4 Temperature Self-Regulation of Hydrogel Microvalve for Thermostatic Control 277
- 13.3.5 Temperature Self-Regulation with Hydrogel Microvalve for Cell Culture 279
- 13.4 Smart Microvalve-in-a-Chip with Ultrasensitivity for Real-Time Detection 281
- 13.4.1 Concept of the Microchip Incorporated with Pb<sup>2+</sup>-Responsive Microgel for Real-Time Online Detection of Trace Pb<sup>2+</sup> 281
- 13.4.2 Sensitivity of the Pb<sup>2+</sup> Detection Platform 283

13.4.3	Selectivity and Repeatability of the Pb <sup>2+</sup> Detection Platform	284
13.4.4	Setup of Pb <sup>2+</sup> Detection System for Real-Time Online Detection of Pb <sup>2+</sup> in Tap Water for Pollution Warning	287
13.4.5	Setup of Pb <sup>2+</sup> Detection System for Real-Time Online Detection of Pb <sup>2+</sup> in Wastewater from a Model Industrial Factory for Pollution Warning and Terminating	289
13.5	Summary	289
	References	289
<b>14</b>	<b>Summary and Perspective</b>	<b>295</b>
14.1	Summary	295
14.2	Perspective	295
	References	297
	<b>Index</b>	<b>299</b>

## Preface

Microfluidics, or the so-called lab-on-a-chip, has emerged as a distinct new technology since the beginning of the 1990s. The dimensions of the microfluidic channels and components are tens to hundreds of micrometers. The microfluidic devices can be used to flexibly manipulate the flow of microvolume fluids in microchannels, which are considered putting the lab on a chip. Due to the trend of miniaturization and integration of modern scientific and technological development, microfluidic technology has been widely concerned and valued by the international scientific and industrial communities. Since microfluidic technology can accurately manipulate small-volume fluids, it is rapidly extending from the original analytical chemistry platform for microanalysis and microdetection to high-throughput drug screening, micromixing, microreaction, microseparation, and so on. Due to its excellent ability to control fluid interfaces as well as excellent heat and mass transfer performances, microfluidic technology has become a novel and promising material preparation technology platform. Microfluidic technology has emerged in the construction of precisely controllable microstructured new functional materials with high performances and especially shows incomparable creativity and superiority compared with traditional technology in the design and preparation of some new functional materials with high added values.

This book, entitled *Microfluidics for Advanced Functional Polymeric Materials*, comprehensively and systematically treats modern understanding of the microfluidic technique and its great power in controllable fabrication of advanced functional polymeric materials. The contents range from the design and fabrication of microfluidic devices, the fundamentals and strategies for controllable microfluidic generation of multiphase liquid systems (e.g., discrete multiple emulsions and continuous laminar multiflow systems), and the use of these liquid systems with elaborate combination of their structures and compositions for controllable fabrication of advanced functional polymeric materials (e.g., solid microparticles, porous microparticles, hollow microcapsules, core-shell microcapsules, hole-shell microcapsules, multicompartmental microcapsules, microfibers, in-chip membranes, and microvalves). All the chapters together clearly describe the design concepts and fabrication strategies of advanced functional polymeric materials with microfluidics by combining the structures with the compositions of multiphase liquid systems to achieve advanced and novel functions. Vivid schematics and illustrations throughout

the book enhance the accessibility to the relevant theory and technologies. This book aims to be a definitive reference book for a wide general readership including chemists, chemical engineers, materials researchers, pharmaceutical scientists, biomedical researchers, and students in the related fields.

The book is composed of 14 chapters. In Chapter 1, a brief introduction of the superiority and potential of microfluidics in the construction of microscale phase interfaces and preparation of novel functional materials are briefly introduced. In Chapters 2 and 3, microfluidic strategies for shear-induced and wetting-induced generations of controllable multiple emulsions are introduced, respectively. In Chapters 4–6, microfluidic strategies for controllable fabrication of monodisperse hydrogel microparticles, porous microparticles, and hierarchical porous microparticles are introduced, respectively. In Chapters 7 and 8, microfluidic strategies for controllable fabrication of monodisperse hollow microcapsules and core–shell microcapsules with an oil core and a stimuli-responsive hydrogel shell are introduced, respectively. In Chapter 9, the microfluidic strategies for fabrication of controllable hole–shell microparticles from double emulsions are introduced. In Chapter 10, a microfluidic strategy for template synthesis of multicompartmental microparticles, with accurate control over the structures of their inner compartments and the encapsulation characteristics of their loaded contents, is introduced. In Chapter 11, simple and versatile microfluidic strategies for controllable fabrication of functional microfibers with tubular, peapod-like, and spindle-knot-like internals are introduced. In Chapter 12, a microfluidic strategy for *in situ* fabrication of nanogel-containing smart membranes in the microchannel of microchips is introduced. In Chapter 13, fabrication and performance of microchips incorporated with smart hydrogel microvalves for thermostatic control and trace analytes detection are introduced. Finally, perspectives on the microfluidic fabrication of advanced functional polymeric materials are given in Chapter 14.

The authors' group at Sichuan University (group website: [http://teacher.scu.edu.cn/ftp\\_teacher0/cly/](http://teacher.scu.edu.cn/ftp_teacher0/cly/)) has been devoted to the microfluidic fabrications of polymeric functional materials since 2006. In the past decade, they have made significant contributions to the development of this field. Most of the contents in this book are the fresh achievements of the authors' group on advanced functional materials fabricated with microfluidics. Prof. Liang-Yin Chu wrote Chapters 1, 4, 5, 7, 12, and 14, and Prof. Wei Wang wrote Chapters 2, 3, 6, 8–11, and 13. The authors are very grateful to Prof. David A. Weitz at Harvard University who helped the authors a lot to carry out investigations in the field of microfluidics. The authors would like to thank all the current and former group members who contributed to the investigations on microfluidics, especially Prof. Rui Xie, Prof. Xiao-Jie Ju, Prof. Zhuang Liu, Dr Li Liu, Dr Nan-Nan Deng, Dr Mao-Jie Zhang, Dr Zhi-Jun Meng, Dr Ya-Lan Yu, Dr Jie Wei, Dr Lei Zhang, Dr Chuan-Lin Mou, Dr Li-Li Yue, Dr Gang Chen, Dr Ying-Mei Liu, Dr Hai-Rong Yu, Dr Xiao-Heng He, Dr Ming-Yue Jiang, Dr Shuo Lin, Dr Fang Wu, Dr Xiao-Yi Zou, Jian Sun, Hao Zhang, Ping-Wei Ren, Jian-Ping Yang, Shuo-Wei Pi, Xi Lin, Guo-Qing Wen, Yi-Meng Sun, Chao Yang, Wei-Chao Zheng, Mei Yuan, Xiu-Lan Yang, and Ming Li, for their creative researches on microfluidics. The authors gratefully acknowledge all the professors, friends, and colleagues who helped the authors' group



to carry out investigations on microfluidics, and thank all the organizations who financially supported the authors' group for the continuous study in the field of microfluidics.

Finally, the authors would like to acknowledge the kind help of Dr Lifen Yang and the editorial staff at Wiley during the preparation and publication of this book.

September 2016

*Liang-Yin Chu*  
*Sichuan University Chengdu, China*



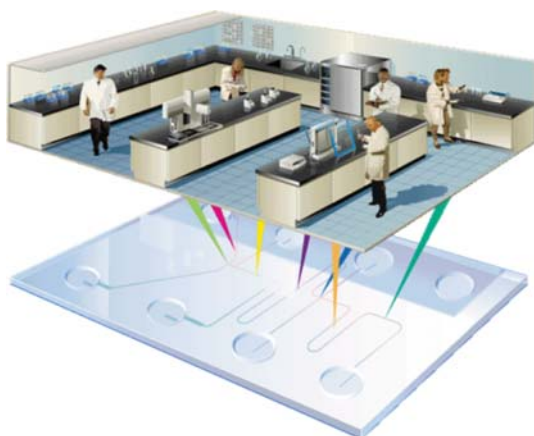
## 1

## Introduction

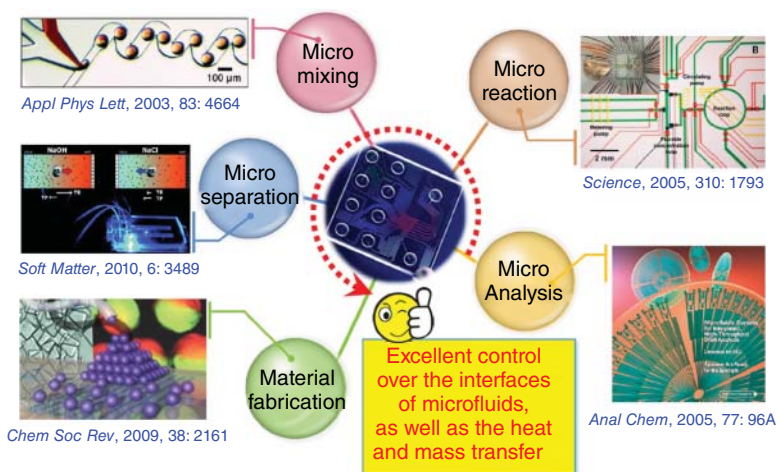
### 1.1 Microfluidics and Its Superiority in Controllable Fabrication of Functional Materials

Microfluidics, or the so-called lab-on-a-chip, has emerged as a distinct new technology since the beginning of 1990s [1]. The dimensions of the microfluidic channels and components are tens to hundreds of micrometers. The microfluidic devices can be used to flexibly manipulate the flow of microvolume fluids in microchannels, which are considered putting the lab on a chip (Figure 1.1). Due to the trend of miniaturization and integration of modern scientific and technological civilization development, microfluidic technology has been widely concerned and valued by the international scientific and industrial communities. In 2006, *Nature* magazine published a special issue on the topic of “*Insight: lab on a chip*,” including seven related review papers [1, 2], where the editorial says that it might have the potential to become “a technology for this century.” In 2010, *Chemical Society Reviews* published a special issue on the topic of “*From microfluidic applications to nanofluidic phenomena*,” including 20 related review papers [3], which shows the promising momentum of development of microfluidic technologies. Since microfluidic technology can accurately manipulate small-volume fluids, it is rapidly extending from the original analytical chemistry platform for microanalysis and microdetection, to high-throughput drug screening, micromixing, microreaction, microseparation, and so on. Due to its excellent ability to control fluid interfaces as well as excellent heat and mass transfer performances, microfluidic technology has become a novel and promising material preparation technology platform (Figure 1.2). Microfluidic technology has emerged in the construction of precisely controllable microstructured new functional materials with high performances, such as microcapsules and microspheres, membranes in microchannels, and superfine fiber materials, and especially shows incomparable creativity and superiority compared with traditional technology in design and preparation of some new functional materials with high added values [4–35].

To sum up, stable phase interface structures of immiscible liquid phase systems that are constructed by the microfluidic technology can be mainly divided into two systems [4]: one is the emulsion droplet system with closed liquid–liquid interfaces, and the second is the laminar flow system with closed

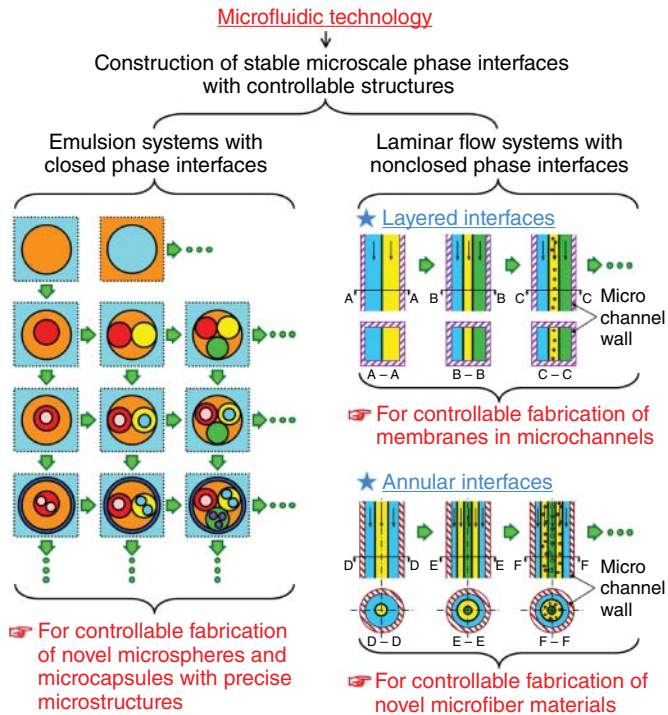


**Figure 1.1** Microfluidics: putting the lab on a chip.



**Figure 1.2** Microfluidic technology is becoming a novel technology platform for materials preparation because of its excellent control over the microfluid interfaces as well as the heat and mass transfer.

liquid–liquid interfaces (Figure 1.3). These two microfluidic-constructed stable phase interface structure systems can be used to prepare three categories of high-performance functional materials with accurate and controllable microstructures as follows [4–35]: (i) controllable fabrication of novel microspheres and microcapsules with precise microstructures by using emulsion droplet systems with closed phase interfaces as templates [4, 5, 7–32]; (ii) controllable fabrication of membranes in microchannels by using laminar flow systems with nonclosed layered phase interfaces [6, 33, 36, 37]; and (iii) controllable fabrication of novel microfiber materials by using laminar flow systems with nonclosed annular phase interfaces [4, 10, 34, 38]. As illustrated in Figure 1.3, microfluidic technology shows superior controllability and great potential in the construction of these three kinds of functional materials, and



**Figure 1.3** The system diagram of microfluidic method for the construction of stable microscale phase interfaces and for controllable preparation of novel functional materials.

can play its unique advantages in controllable construction of new functional materials with new structures, new functions, and high-performance features.

## 1.2 Microfluidic Fabrication of Microspheres and Microcapsules from Microscale Closed Liquid–Liquid Interfaces

Due to the small size and controllable internal structure, microspheres and microcapsules can be used as microcarriers, microreactors, microseparators, and microstructural units in drug delivery, substance encapsulation, chemical catalysis, biochemical separation, artificial cells, and enzyme immobilization, and have very broad application prospects. Microspheres and microcapsules are generally fabricated by using emulsion droplets with stable closed liquid–liquid interfaces (e.g., single water-in-oil (W/O) or oil-in-water (O/W) emulsions, W/O/W or O/W/O double emulsions, or even more complicated multiple emulsions) as templates, through subsequent polymerization, cross-linking, solvent evaporation, curing, and assembling in emulsion droplets or at interfaces. Traditional methods for the preparation of emulsion droplets are mainly achieved by mechanical stirring or fluid shear; thus, the sizes and the internal

structures of the droplets and the resultant template-fabricated microspheres and microcapsules are difficult to be controlled precisely, which greatly affect the performances and applications of the microspheres and microcapsules.

Microfluidic technology, which can generate emulsion droplets by emulsifying disperse phase to continuous phase through microchannels with co-flow, flow-focusing, or T-junction geometries, can achieve continuous and precise control of the microstructures of emulsion droplets, exhibiting significant superiority in the fabrication of microspheres and microcapsules with controllable size distributions and microstructures.

Researchers from all over the world have made a lot of important progress in the use of microfluidics to construct microscale closed liquid–liquid interfaces and then fabricate monodisperse microspheres and microcapsules [4, 5, 7–32]. In the preparations of microspheres and microcapsules with microfluidic approaches, most of them are focused on the use of microfluidic-generated W/O or O/W single emulsions (as shown in the first row in the upper left corner of Figure 1.3) as templates for preparing monodisperse microspheres, or the use of W/O/W or O/W/O double emulsions (as shown in the second row in the first column of the upper left corner of Figure 1.3) as templates for preparing monodisperse core–shell microcapsules. Some studies have also attempted to prepare some materials with new structures such as multicore microspheres, Janus microspheres, and nonspherical particles by microfluidic technology.

The authors' group controllably constructed multiple emulsion systems with complex microscale multiphase multicomponent liquid–liquid interfaces by building series and parallel microchannels [31]. These emulsions are used as templates for controllably preparing multiphase multicomponent microspheres and microcapsules for the encapsulation of substances [29], as well as new multifunctional microspheres and microcapsules with complex structures [28, 30].

### 1.3 Microfluidic Fabrication of Membranes in Microchannels from Microscale Nonclosed Layered Laminar Interfaces

Because of the excellent performances in catalysis, separations, purifications, analysis and detection, controlled release, emulsification, and so on, functional membrane materials are considered as one of the important supporting technologies for sustainable development. If the combination of membrane materials and microfluidic technology is obtained, it will play the synergy of the two to achieve the integration of functional materials and components. In this way, it can not only promote the application of membrane materials in microseparation and microanalysis but also provide new catalysis- or reaction-separation coupling technologies for microchemical or microreaction processes, showing very broad application prospects [6]. Therefore, as a new technology platform, membrane-in-microchannel technology is increasingly subject to different disciplines of international attention [6].

In a co-flow microchannel, when immiscible multiphase fluids flows into the same microchannel, stable layered laminar flow patterns can be formed through

microfluidic laminar flow technology [36] (“Layered interfaces” in Figure 1.3). In each phase, the fluid can maintain its flow pattern unchanged; chemical reactions such as polymerization and cross-linking only occur at the liquid–liquid interfaces, forming monolayer or multilayer parallel ultrathin membranes in the microchannels.

The microchannels can be divided into several independent channels by the membranes in microchannels. Due to the selective permeability or adsorption ability of functional membrane materials, selective separation, extraction, detection, and analysis can be realized with the membranes in the microchannels. Catalysts can also be effectively deposited on the membrane surfaces, thereby increasing the specific surface area of the catalytic material within a microchannel, to accelerate the rate of catalytic reaction in the microchannel. In addition, environmental stimuli-responsive smart membranes, which can regulate the effective membrane pore size and permeability in response to the change in physical or chemical signals in the environment, show incomparable superiority over traditional membranes [39]. If the smart membranes can be combined with microfluidics, it will undoubtedly provide efficient technology platform for the intensification of microseparation, microanalysis and detection, microreaction processed, and the enhancement of membrane performances.

Since the fabrication processes of membranes in microchannels are different from traditional membrane preparation processes, so far there are only a few reports on the fabrications of membranes with limited materials such as polyamide and chitosan in microchannels by using microfluidic laminar flow technology [6, 33, 36, 37, 40, 41].

## 1.4 Microfluidic Fabrication of Microfiber Materials from Microscale Nonclosed Annular Laminar Interfaces

Microfiber materials have a wide range of applications in optoelectronics, biomedicine, chemical industry, light industry, and other fields, wherein the hollow fiber membranes play an important role in the chemical separation processes. Currently, the preparations of microfiber materials are mainly achieved by using melt spinning method, electrospinning method, and other methods, while these methods are still difficult to achieve precise control of the microstructures of microfiber materials or impart multifunctional characteristics. Therefore, it is still necessary to seek new preparation processes and methods for the preparation of microfiber materials, and the microfluidic laminar flow technology is a very promising new method.

With microfluidic laminar flow technology, stable annular laminar flow patterns of immiscible multiphase fluids can be formed in the microchannels [4] (“Annular interfaces” in Figure 1.3). With these stable annular multiphase laminar interface systems as templates, microfiber materials including linear solid microfibers, hollow tubular microfibers, and core–shell composite microfibers can be fabricated by reaction or curing at the liquid–liquid interfaces or inside phases.

Since microfluidic technology enables continuous and accurate control over the annular liquid–liquid interfaces of laminar flows, it can provide optimal design of fibrous material synthesis systems. Therefore, compared with traditional spinning techniques, microfluidic technology has significant advantages in precise regulation and design of microfiber microstructures: it improves performances and imparts multifunctional characteristics of microfiber materials [4, 34, 36, 38, 42–47].

Microfluidic technology has been used to successfully prepare calcium alginate, polyvinyl alcohol, poly(lactic-co-glycolic acid), liposomes, chitosan, poly(ether sulfone), and polyacrylonitrile microfibers [34, 38, 42–47], which show excellent flexibility and extraordinary potential in the construction of microscale annular liquid–liquid interfaces and preparation of microfiber materials.

## References

- 1 Whitesides, G.M. (2006) The origins and the future of microfluidics. *Nature*, **442**, 368–373.
- 2 Daw, R. and Finkelstein, J. (2006) Insight: lab on a chip. *Nature*, **442**, 367–418.
- 3 van den Berg, A., Craighead, H.G., and Yang, P. (2010) From microfluidic applications to nanofluidic phenomena. *Chem. Soc. Rev.*, **39**, 899–1217.
- 4 Atencia, J. and Beebe, D.J. (2005) Controlled microfluidic interfaces. *Nature*, **437**, 648–655.
- 5 Joanicot, M. and Ajdari, A. (2005) Applied physics – droplet control for microfluidics. *Science*, **309**, 887–888.
- 6 de Jong, J., Lammertink, R.G.H., and Wessling, M. (2006) Membranes and microfluidics: a review. *Lab Chip*, **6**, 1125–1139.
- 7 Utada, A.S., Chu, L.Y., Fernandez-Nieves, A., Link, D.R., Holtze, C., and Weitz, D.A. (2007) Dripping, jetting, drops, and wetting: the magic of microfluidics. *MRS Bull.*, **32**, 702–708.
- 8 Shah, R.K., Shum, H.C., Rowat, A.C., Lee, D., Agresti, J.J., Utada, A.S., Chu, L.Y., Kim, J.W., Fernandez-Nieves, A., Martinez, C.J., and Weitz, D.A. (2008) Designer emulsions using microfluidics. *Mater. Today*, **11**, 18–27.
- 9 Dendukuri, D. and Doyle, P.S. (2009) The synthesis and assembly of polymeric microparticles using microfluidics. *Adv. Mater.*, **21**, 4071–4086.
- 10 Tumarkin, E. and Kumacheva, E. (2009) Microfluidic generation of microgels from synthetic and natural polymers. *Chem. Soc. Rev.*, **38**, 2161–2168.
- 11 Theberge, A., Courtois, F., Schaerli, Y., Fischlechner, M., Abell, C., Hollfelder, F., and Huck, W. (2010) Microdroplets in microfluidics: an evolving platform for discoveries in chemistry and biology. *Angew. Chem. Int. Ed.*, **49**, 5846–5868.
- 12 Marre, S. and Jensen, K.F. (2010) Synthesis of micro and nanostructures in microfluidic systems. *Chem. Soc. Rev.*, **39**, 1183–1202.
- 13 Chu, L.Y., Utada, A.S., Shah, R.K., Kim, J.W., and Weitz, D.A. (2007) Controllable monodisperse multiple emulsions. *Angew. Chem. Int. Ed.*, **46**, 8970–8974.



- 14 Chu, L.Y., Kim, J.W., Shah, R.K., and Weitz, D.A. (2007) Monodisperse thermo-responsive microgels with tunable volume-phase transition kinetics. *Adv. Funct. Mater.*, **17**, 3499–3504.
- 15 Wang, W., Liu, L., Ju, X.J., Zerrouki, D., Xie, R., Yang, L.H., and Chu, L.Y. (2009) A novel thermo-induced self-bursting microcapsule with magnetic-targeting property. *ChemPhysChem*, **10**, 2405–2409.
- 16 Zhou, M.Y., Xie, R., Ju, X.J., Zhao, Z.L., and Chu, L.Y. (2009) Flow characteristics of thermo-responsive microspheres in microchannel during the phase transition. *AIChE J.*, **55**, 1559–1568.
- 17 Liu, L., Yang, J.P., Ju, X.J., Xie, R., Yang, L.H., Liang, B., and Chu, L.Y. (2009) Microfluidic preparation of monodisperse ethyl cellulose hollow microcapsules with non-toxic solvent. *J. Colloid Interface Sci.*, **336**, 100–106.
- 18 Zhang, H., Ju, X.J., Xie, R., Cheng, C.J., Ren, P.W., and Chu, L.Y. (2009) A microfluidic approach to fabricate monodisperse hollow or porous poly(HEMA-MMA) microspheres using single emulsions as templates. *J. Colloid Interface Sci.*, **336**, 235–243.
- 19 Liu, L., Wang, W., Ju, X.J., Xie, R., and Chu, L.Y. (2010) Smart thermo-triggered squirting capsules for nanoparticle delivery. *Soft Matter*, **6**, 3759–3763.
- 20 Ren, P.W., Ju, X.J., Xie, R., and Chu, L.Y. (2010) Monodisperse alginate microcapsules with oil core generated from a microfluidic device. *J. Colloid Interface Sci.*, **343**, 392–395.
- 21 Pi, S.W., Ju, X.J., Wu, H.G., Xie, R., and Chu, L.Y. (2010) Smart responsive microcapsules capable of recognizing heavy metal ions. *J. Colloid Interface Sci.*, **349**, 512–518.
- 22 Yu, Y.L., Xie, R., Zhang, M.J., Li, P.F., Yang, L.H., Ju, X.J., and Chu, L.Y. (2010) Monodisperse microspheres with poly(*N*-isopropylacrylamide) core and poly(2-hydroxyethyl methacrylate) shell. *J. Colloid Interface Sci.*, **346**, 361–369.
- 23 Wei, J., Ju, X.J., Xie, R., Mou, C.L., Lin, X., and Chu, L.Y. (2011) Novel cationic pH-responsive poly(*N,N*-dimethylaminoethyl methacrylate) microcapsules prepared by a microfluidic technique. *J. Colloid Interface Sci.*, **357**, 101–108.
- 24 Liu, L., Yang, J.P., Ju, X.J., Xie, R., Liu, Y.M., Wang, W., Zhang, J.J., Niu, C.H., and Chu, L.Y. (2011) Monodisperse core-shell chitosan microcapsules for pH-responsive burst release of hydrophobic drugs. *Soft Matter*, **7**, 4821–4827.
- 25 Liu, L., Wu, F., Ju, X.J., Xie, R., Wang, W., Niu, C.H., and Chu, L.Y. (2013) Preparation of monodisperse calcium alginate microcapsules via internal gelation in microfluidic-generated double emulsions. *J. Colloid Interface Sci.*, **404**, 85–90.
- 26 Zhang, M.J., Wang, W., Xie, R., Ju, X.J., Liu, L., Gu, Y.Y., and Chu, L.Y. (2013) Microfluidic fabrication of monodisperse microcapsules for glucose-response at physiological temperature. *Soft Matter*, **9**, 4150–4159.
- 27 Wang, W., Yao, C., Zhang, M.J., Ju, X.J., Xie, R., and Chu, L.Y. (2013) Thermo-driven microcrawlers fabricated via a microfluidic approach. *J. Phys. D: Appl. Phys.* (Special Issue on Microfluidics), **46**, 114007.

- 28 Wang, W., Zhang, M.J., Xie, R., Ju, X.J., Yang, C., Mou, C.L., Weitz, D.A., and Chu, L.Y. (2013) Hole-shell microparticles from controllably evolved double emulsions. *Angew. Chem. Int. Ed.*, **52**, 8084–8087.
- 29 Wang, W., Luo, T., Ju, X.J., Xie, R., Liu, L., and Chu, L.Y. (2012) Microfluidic preparation of multicompartment microcapsules for isolated co-encapsulation and controlled release of diverse components. *Int. J. Nonlinear Sci. Numer. Simul.* (Special Issue on Microfluidics), **13**, 325–332.
- 30 Liu, Y.M., Wang, W., Zheng, W.C., Ju, X.J., Xie, R., Zerrouki, D., Deng, N.N., and Chu, L.Y. (2013) Hydrogel-based micro-actuators with remote-controlled locomotion and fast  $\text{Pb}^{2+}$ -response for micromanipulation. *ACS Appl. Mater. Interfaces*, **5**, 7219–7226.
- 31 Wang, W., Xie, R., Ju, X.J., Luo, T., Liu, L., Weitz, D.A., and Chu, L.Y. (2011) Controllable microfluidic production of multicomponent multiple emulsions. *Lab Chip*, **11**, 1587–1592.
- 32 Wang, W., Zhang, M.J., and Chu, L.Y. (2014) Functional polymeric microparticles engineered from controllable microfluidic emulsions. *Acc. Chem. Res.*, **47**, 373–384.
- 33 Sun, Y.M., Wang, W., Wei, Y.Y., Deng, N.N., Liu, Z., Ju, X.J., Xie, R., and Chu, L.Y. (2014) *In situ* fabrication of temperature- and ethanol-responsive smart membrane in microchip. *Lab Chip*, **14**, 2418–2427.
- 34 He, X.H., Wang, W., Liu, Y.M., Jiang, M., Wu, F., Deng, K., Liu, Z., Ju, X.J., Xie, R., and Chu, L.Y. (2015) Microfluidic fabrication of bio-inspired microfibers with controllable magnetic spindle-knots for 3D assembly and water collection. *ACS Appl. Mater. Interfaces*, **7**, 17471–17481.
- 35 Lin, S., Wang, W., Ju, X.J., Xie, R., Liu, Z., Yu, H.R., Zhang, C., and Chu, L.Y. (2016) Ultrasensitive microchip based on smart microgel for real-time on-line detection of trace threat analytes. *Proc. Natl. Acad. Sci. U. S. A.*, **113**, 2023–2028.
- 36 Kenis, P.J.A., Ismagilov, R.F., and Whitesides, G.M. (1999) Microfabrication inside capillaries using multiphase laminar flow patterning. *Science*, **285**, 83–85.
- 37 Hisamoto, H., Shimizu, Y., Uchiyama, K., Tokeshi, M., Kikutani, Y., Hibara, A., and Kitamori, T. (2003) Chemicofunctional membrane for integrated chemical processes on a microchip. *Anal. Chem.*, **75**, 350–354.
- 38 Lan, W.J., Li, S.W., Lu, Y.C., Xu, J.H., and Luo, G.S. (2009) Controllable preparation of microscale tubes with multiphase co-laminar flow in a double co-axial microdevice. *Lab Chip*, **9**, 3282–3288.
- 39 Chu, L.Y. (2011) *Smart Membrane Materials and Systems*, Springer, Berlin and Heidelberg.
- 40 Uozumi, Y., Yamada, Y.M.A., Beppu, T., Fukuyama, N., Ueno, M., and Kitamori, T. (2006) Instantaneous carbon–carbon bond formation using a microchannel reactor with a catalytic membrane. *J. Am. Chem. Soc.*, **128**, 15994–15995.
- 41 Luo, X.L., Berlin, D.L., Betz, J., Payne, G.F., Bentley, W.E., and Rubloff, G.W. (2010) In situ generation of pH gradients in microfluidic devices for biofabrication of freestanding, semi-permeable chitosan membranes. *Lab Chip*, **10**, 59–65.

- 42 Jeong, W., Kim, J., Kim, S., Lee, S., Mensing, G., and Beebe, D.J. (2004) Hydrodynamic microfabrication via “on the fly” photopolymerization of microscale fibers and tubes. *Lab Chip*, **4**, 576–580.
- 43 Dittrich, P.S., Heule, M., Renaud, P., and Manz, A. (2006) On-chip extrusion of lipid vesicles and tubes through microsized apertures. *Lab Chip*, **6**, 488–493.
- 44 Shin, S.J., Park, J.Y., Lee, J.Y., Park, H., Park, Y.D., Lee, K.B., Whang, C.M., and Lee, S.H. (2007) “On the fly” continuous generation of alginate fibers using a microfluidic device. *Langmuir*, **23**, 9104–9108.
- 45 Hwang, C.M., Khademhosseini, A., Park, Y., Sun, K., and Lee, S.H. (2008) Microfluidic chip-based fabrication of PLGA microfiber scaffolds for tissue engineering. *Langmuir*, **24**, 6845–6851.
- 46 Puigmarti-Luis, J., Schaffhauser, D., Burg, B.R., and Dittrich, P.S. (2010) A microfluidic approach for the formation of conductive nanowires and hollow hybrid structures. *Adv. Mater.*, **22**, 2255–2259.
- 47 Lan, W.J., Li, S.W., Xu, J.H., and Luo, G.S. (2012) Controllable synthesis of microscale titania fibers and tubes using co-laminar micro-flows. *Chem. Eng. J.*, **181–182**, 828–833.



## 2

## Shear-Induced Generation of Controllable Multiple Emulsions in Microfluidic Devices

### 2.1 Introduction

Multiple emulsions are complex nested liquid systems, containing liquid droplets of decreasing sizes placed one inside another. They are widely used as encapsulation systems in myriad applications, including drug delivery [1–3], foods [4, 5], cosmetics [6, 7], chemical separations [8, 9], and as templates for syntheses of microspheres and microcapsules [10–16]. Accurate control of the size monodispersity and internal structure of multiple emulsions are critical for their versatility because these features enable precise manipulation of the loading levels and the release and transport kinetics of the encapsulated substances [10–21]. Although there have been several reports on the preparation of monodisperse multiple emulsions [13–23], accurate control of both the size and structure of the emulsions remains difficult to achieve, and current techniques are not scalable for fabricating higher order multiple emulsions.

Typically, multiple emulsions can be generated via sequential bulk emulsification using shear. This method usually produces emulsions with polydisperse size. Controlling the shear with constrained geometry [24–28], porous membranes [17, 21], or microchannels [18] can produce emulsions with nearly monodisperse size; these can then themselves be emulsified to produce multiple emulsions. However, although the volume fractions of both the initial and final emulsions can be manipulated [17, 18, 21, 24–27], accurate control over the number of inner droplets in multiple emulsions remains difficult by using these techniques. However, there are many cases where control of the inner droplet number is more important than control of their volume fraction. For example, with precise control of the number and size of inner droplets, transport kinetics of encapsulated substances in the emulsions can be precisely manipulated. Control of the inner droplet number is also critical for engineering colloidal assemblies to produce nonspherical particles [29]. Moreover, it is still challenging for co-encapsulation of droplets containing distinct contents in multiple emulsions, with accurate control of the number, ratio, and size of different inner droplets within each level. With such control on the structure of multicomponent multiple emulsions, these emulsions can offer advanced platforms for design of more complex multicompartiment materials and provide synergistic delivery systems or chemical microreactors for incompatible actives or chemicals with versatile encapsulation and flexible mass-transfer kinetics.

Microfluidic techniques provide an alternate route to generate monodisperse multiple emulsions. Cascading two T-junction geometries [19, 23] can generate monodisperse double emulsions with some control over the number and size of inner droplets. However, the microfluidic device with standard two-dimensional (2D) microchannels requires precise, localized modification of the microchannel wettability for producing multiple emulsions. This ultimately restricts their utility. By contrast, coaxial flow-focusing [13, 14, 16] geometries relax the constraints for wettability modification, but are still limited in the range of fluids that can be used and in the accurate control afforded over the number and size of inner droplets. Moreover, co-encapsulation of multicomponent droplets in multiple emulsions with precise control of their number, ratio, and size within each level, cannot be easily achieved with these devices. In addition, current microfluidic approaches cannot be scaled up for producing higher order multiple emulsions, such as triple emulsions.

The authors' group developed a highly scalable and versatile microfluidic strategy for controllable shear-induced generation of monodisperse multiple emulsions. In this chapter, the microfluidic devices for controllable emulsion generation and the controllable microfluidic fabrication of single emulsions, double emulsions, triple emulsions, and multicomponent multiple emulsions with even more complex structures are introduced.

## 2.2 Microfluidic Strategy for Shear-Induced Generation of Controllable Emulsion Droplets

Usually, microfluidic devices for generating emulsion droplets consist of droplet-making units and connecting units. The droplet-making units usually contain microchannels with co-flow (Figure 2.1a) [31], flow-focusing (Figure 2.1b) [32], and T-junction (Figure 2.1c) [19] geometries for generating emulsion droplets, while the connecting units usually contain microchannels for manipulating the generated droplets.

Typically, the microchannels in microfluidic device can be constructed by coaxially inserting cylinder glass capillaries into square glass tubes. The inner microchannel of the inserted capillary and the interstice space between the inserted capillary and square tube create three-dimensional (3D) microchannels for flowing different fluids. After fixing such assembly structures of glass capillaries and tubes on glass plates, microfluidic device with microchannels for emulsion generation can be fabricated [33, 34]. The glass-capillary microfluidic device can be used for emulsion generation without microchannel surface modification due to their 3D microchannel structure, but the fabrication process of these devices requires troublesome manual assembling. Alternatively, based on soft lithography, 2D microchannels can be etched on polydimethylsiloxane (PDMS) plates for fabrication of microfluidic devices [35]. These PDMS microfluidic devices allow flexible construction of microchannel networks for generation and manipulation of droplets. Meanwhile, their fabrication process based on the well-established soft lithography technique allows massive production of the devices. However, these PDMS devices require troublesome